Review Article

Functional, nutritional and medicinal potential of banana peel

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Abstract

Banana, botanically called Musa spp. is among the most cultivated and consumed fruits, which generates a huge amount of waste (peel) annually. The current review was to combine the activities and beneficial aspects of banana peel. The organic composition of banana peel comprising lipids, fibers, carbohydrates and protein which results in presence of several bioactive molecules. The literature revealed that these bioactive compounds including alkaloids, flavonoids, phenolics, steroids (tannic acid, catechol, β -sitosterol, ferulic acid) help to perform different biological activities and cures antitumor, antiparasitic, antibacterial, antifungal, antiaging, antioxidant and antiviral activities. The mechanism of its bioactive compounds helps to cure infections and diseases. The study states that banana peel is a valuable byproduct which has several benefits and it can be used in different industries like pharmaceutical, cosmetics, food, lather, biodiesel and bioethanol.

Keywords: Amino acids; Anti-inflammatory; Antimicrobial; Antioxidant; Dermatological effect; Phytochemicals

Introduction

Banana (Musa spp.) is one of the most commonly eaten fruits with considerable nutritional value [1]. In tropical and subtropical zone, this crop is widely grown and consumed. Most of this fruit is consumed fresh, but it can also be processed into a variety of products at large scale, including dried fruit, chips, smoothies, ice cream, bread, flour, wine, and ingredients for functional cuisine [2-4]. Literature indicated, Southern Asian tropical regions are where bananas were first domesticated; today, 130 nations grow bananas. Banana production has significantly expanded globally over the last 20 years. The average annual output of bananas worldwide has increased from 69 million tons in 2000-2002 to 116 million tons in 2017-2019, in which India stands first with a production of 32 million metric tons annually (Fig. 1) [5]. Pakistan has produced 154,800 tons of bananas from 34,800 hectares on average. Sindh province of Pakistan has the major share with 87 % of total area followed by Khyber Pakhtunkhwa (KPK), Punjab and Balochistan [6].

The genus Musa of the family Musaceae includes a wide range of hybrids. According to published reports, there are almost 30 different varieties of banana present all over the world [7]. Whereas the *Musa* spp. is further diversified into different four sections Rhodochlamys, namely Eumusa, Australimusa and Callimusa. Among these sections, the most widespread section geographically is *Eumusa* and it is followed by Australimusa. The edible class of bananas is solely from Musa accuminata [8]. Pakistan produce different verities of bananas among which 95 % of area is under Basrai variety (Cavendish dwarf), whereas other common verities are William Hybrid, Grand Nine (G-

9), Chinese varieties B-10, W-11 and Pashing [6].

Banana sector has low yield and is less profitable in Pakistan than that of other neighboring banana-growing countries. Pakistan's banana fruit yield is facing several challenges in which the most significant challenge is prevalence of banana pathogen epidemics and the lack of high yielding varieties that are compatible with the changing climate. Conserving infected suckers, lack of healthy disease-free suckers, prevalence of single low yield variety, poor crop management, skills and technology, water scarcity, fertilizers and post-harvest losses are other challenges [9].

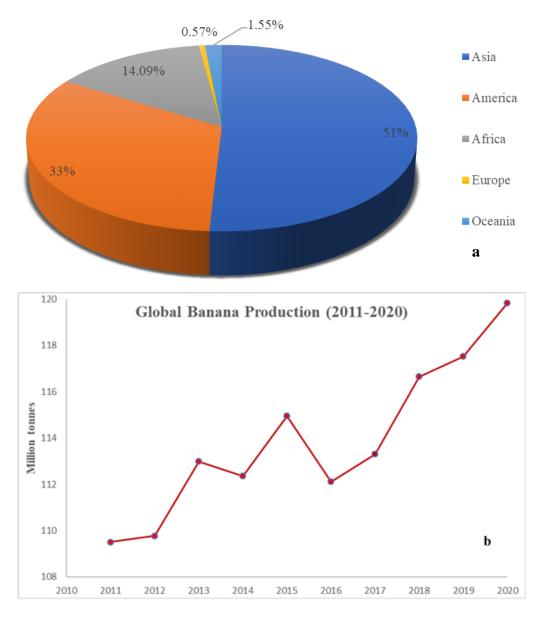


Figure 1: (a) Banana production region-wise (b) Top 10 largest banana-producing countries [5]

Approximately 220 tons of banana deposit produce per hectare comprised chiefly of lignocellulosic material. These deposit wastes are dumped largely into canals, waters

and roadsides origins grave environmental complications [10]. Leaves, peels, psudostems, and stalks all contain a high level of lignocellulose which are the key

deposits of banana produce [11]. The outer coat of banana fruit accounts for up to 35 % of the total mass. Consequently, over 36 million tons of banana peel are produced annually and this is considered to be the potential material for further application. Still, a record percentage of banana peel is disposed of in landfills or with other junk [12]. As we all know that it has very high contents of micronutrients which attracts great attention to its nutritional antioxidant properties, mainly due to the ascorbates, catechin, gallocatechin and dopamine [13, 14].

Additionally, the banana peel also contains various bioactive elements namely terpenoids, alkaloids, flavonoids, tannins and phlobatannins [15]. It has been used as a therapeutic agent for antilithic. antiulcerogenic anti-diarrheal, and hypoglycemics, hypolipidemic and antivenom properties [16-19]. It is said to be beneficial for maintaining very

cholesterol level [20]. Banana peel has a significant amount of carotenoid (64 μ g/g) [21], which acts as a preventive strategy against several tumors, cardiovascular disease, diabetes, and age-related macular degeneration [22]. Banana residue has high phenolic content than avocado, pineapple, papaya, passion fruit, watermelon and melon [23].

Banana peel has high carbon content and can be used as an absorbent to remove various pollutants from savage water [24, 25]. It constitutes hemicelluloses, cellulose and pectin having carboxyl, hydroxyl and amine as a functional group. Rhodamine- B (Rh-B) is said to be among the toxic dyes which cause carcinogenic and neurotoxic effects in humans. Agro-waste has been known for Rh-B removal with great efficiency. Whereas, the above functional group in banana peel has essential value for the binding of Rh-B on bio sorbent (Fig. 2) [26].

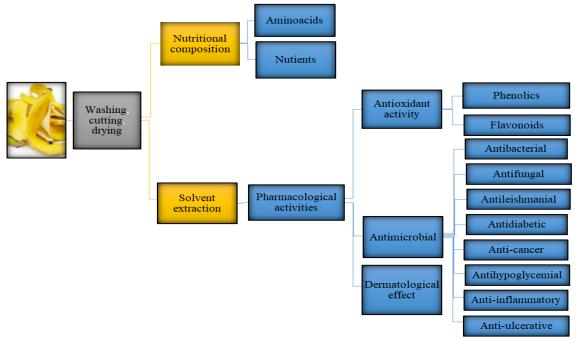


Figure 2: Valorization of banana peel

Nutritional composition of banana peel

Banana peel is a cheap source of starch, crude lipids, proteins, dietary fibers and polyunsaturated fatty acids, especially linoleic and alpha-linolenic acid, pectin, minerals and essential amino acids (Table 1).

It contains amino acids including lysine, leucine, valine, phenylalanine and threonine they are all approved to be higher than Food and Agricultural Organization (FAO) standard except lysine (Table 2) [27].

Table 1: Proximate composition of banana peel and pulp

Contents	Peel	Pulp	Reference
Moisture	75.3±3.2	62.0±2.87	
Non-reducing sugar	3.4±0.19	5.4±0.09	
Protein	5.3±0.02	0.83±0.04	[28-32]
Crude Fat	5.93±0.13	2.58±0.12	[20-32]
Ash	15.29±0.05	1.54±0.07	
Carbohydrates	8.8±0.54	32.5±1.99	

Additionally, banana peel is a good source of cellulose (7.6-9.6 %), lignin (6-12 %), pectin (10-21 %), glucose, galactose, arabinose, rhamnose, xylose and hemicelluloses (6.4-9.4 %) [33]. Micronutrients include calcium, iron, copper, manganese, sodium, potassium,

magnesium, zinc and phosphorus (Values are given below in Table 3) among which mopper, manganese, calcium, magnesium and potassium. Peel has a higher proportion of iron than pulp [34].

Table 2: Amino acids in banana peel and their applications

Amino acids		Banana peel WHO		Application	References	
2 1111		(g/100g)	(g/100g)		References	
	Threonine	2.82	4.7	Substrate for protein synthesis	[35]	
	Valine	2.01	6.6	Involve in muscle growth and energy production	[36]	
	Histidine	4.61	-	Active site of enzymes	[37]	
Tyrosine		1.97	6.0	An essential component for the production of brain chemicals	[38]	
Essential amino	Lysine	7.16	7.0	Substrate for bones and connective tissues	[39]	
acids Methionine		2.52 6	6	Regulate metabolism and innate immune system	[40]	
	Tryptophan	7.39	-	Role in metabolism	[41]	
	Phenylalanine	9.56	9.3	Important in the nervous system	[42]	
	Isoleucine	9.50	5.4	Role in maintaining the immune system	[43]	
	Leucine	10.97	8.6	Stimulate protein synthesis	[44]	
	Asparagine	0	-	Role in nitrogen metabolism in plants	[45]	
Non-	Arginine	11.10	-	Potential role in dentistry	[46]	
essential amino	Serine	8.26	-	Role in plant metabolism and development	[47]	
acids	Glycine	7.32	-	As a biological modifier	[48]	
	Alanine	8.51	-	In dye industry	[49]	
	Cysteine	6.30	-	An important site of protein	[50]	

Table 3: Mineral composition of banana and plantain peel [51]

Minerals	Peel (mg/100g)	Plantain (mg/100g)
Copper	1.35±0.05	0.59±0.83
Iron	5.06±0.07	7.89±0.79
Manganese	10.38±0.04	1.25±0.39
Zinc	11.60±0.03	13.30±0.57

Calcium	17.85±0.25	14.70±0.25
Magnesium	49.32±0.74	45.21±4.36
Sodium	58.16±2.73	76.88±0.89
Potassium	38.22±0.16	26.14±2.68
phosphorus	22.64±0.38	28.95±0.94

Source of bioactive compounds

Recovering bioactive substances from agricultural waste is an effective, affordable, and environmentally friendly strategy to reduce pollution. Numerous uses for these bioactive substances can be found in the food, cosmetics and pharmaceutical industries. Banana peel contains syringic acid, which has the potential to be used to treat abnormalities in glycoprotein components and has an antidiabetic effect in experimental diabetes [52], tannic acid which has been used as a burns treatment agent [53], catechol is used as an intermediary for in lubricating oil and rubber, developer for fur dyes, photographic developer, in polymerization inhibitors and in pharmaceuticals [54], while catechin has resistance of low density lipoprotein (LDL) oxidation, brachial artery dilation increased plasma activity and fat oxidation gallic acid have potential [55], of hepatoprotective effects [56], cinnamic acid has been used as a precursor to the artificial sweetener aspartame by the means of enzyme catalyzed amino group phenylalanine [57], p-coumaric acid has antioxidant characteristics and may lower the chance of stomach cancer [58], while quercetin improves cardiovascular health by improving blood flow [59], ferulic acid is an antioxidant, antimicrobial, antiinflammatory, antiallergic, anticarcinogenic, modulation of enzyme activity, antiviral and vasodilatory actions [60], trans-α carotene, a precursor to vitamin A and trans-β carotene lower the rate of cancer and cardiovascular diseases (CVD) [61]. Violaxanthin is a food colorant while neoxanthin is an intermediary in the generation of the plant hormone abscisic acid [62], cryptoxanthin is used as a food colorant and might reduce the risk of lung cancer [54]. Serotonin may play a role in sensations of happiness and well-being [63], dopamine decreases plasma oxidative stress increases resistance to oxidative modification of low density lipoprotein (LDL) [20] and β-sitosterol able to lower blood cholesterol levels and the risk of benign prostatic hyperplasia [64], campesterol and stigmasterol control the level of cholesterol that is absorbed in human intestines [65], cycloartenol is the primary precursor in the synthesis of steroids (Table 4) [66].

Table 4: Bioactive compounds extracted from banana peel

Bioactive compounds	Sub-types Sub-types	References
	Gallocatechin	
	Glutathione	
Flavan-3-ols	Norepinephrine	[67, 60]
Flavali-3-018	Naringin	[67-69]
	Salsolinol	
	L-Dopa	
	Rutin	
	Quercetin-deoxyhexose hexoside	[59, 69, 70]
Flavanol	Myricetin-deoxyhexose-hexoside	[59, 09, 70]
Plavalioi	Laricitrin-3-rutinoside	
	Syringetin-3-rutinoside	
	Kaempferol-7-rutinoside	
Triterpenic	31-Norcyclolaudenone	[71]
Titterpenic	Cycloeucalenone	[/1]
	Trans-α-carotene,	
Carotenoids	Trans-β-carotene	[54, 62, 72]
	α-Cryptoxanthin	

	β-Cryptoxanthin	
	Violaxanthin	
	Auroxanthin	
	Neoxanthin	
	β-Sitosterol	
Sterols	Stigmasterol	[64, 65]
	Campesterol	
	Cycloeucalenol	
Triterpenes	Cycloartenol	[66, 71]
_	31-Norcyclolaudenone	
	3-Epicycloeucalenol	
Cycloartane-type	3-Epicyclomusalenol	[71, 73]
triterpenes	24-Methylenepollinastanone	
	28-Norcyclomusalenone	
	Methyl palmitate	
	Methyl Linoleate	
	Methyl petroselinate	
Fatty acids	Methyl stearate	[74]
Fatty acids	5-Cholene, 3, 24-dihydroxy	
	5-Dihydroergosterol	
	4,8,13-Duvatriene-1, 3-diol	
	Henicosyl formate	
	Serotonin	
Active amines	Tyramine	[63, 75, 76]
	Norepinephrine	
	Putrescine	
Polyamines	Spermidine	[77, 78]
	Spermine	

Antioxidant potential of banana peel

Antioxidants are substances that neutralize free radicals and can be obtained from food. antioxidants play pharmacologically role in improving male reproductive health, reducing insulin resistance and lowering the risk of type II diabetes [79, 80]. Antioxidants promote normal biological functions including healthy cell proliferation, immunological support and molecular degeneration prevention, including the prevention of premature ageing [81]. The banana should be regarded as a useful dietary source against diseases and an excellent source of natural antioxidants for foods. These compounds predominate in the majority of the tissues in banana peel [82]. The banana peel extract was found to contain gallocatechin which demonstrated significant antioxidant activity. Shinichi et al. [83] revealed in her study that peel contained higher gallocatechin (158 mg/100g DW) than pulp (29.6 mg/100g DW). It was also more potent than the banana pulp extract against lipid autoxidation. Catechins show strong antioxidant effects against lipid peroxidation [84] and protective effects against diseases such as cancer and CVD [85]. Antioxidant capacity depends on different solvents such as acetone, acetyl acetate, chloroform, ethanol, methanol and n-hexane by several antioxidant potential determining assays including diphenyle-1-picrylhydrazyl assay (DPPH), ferric reducing antioxidant power (FRAP), metal chelating activity, 2,2'-azinobis (3-ethylebenzothiazoline-6-sulfunic acid assay (ABTS), superoxide anion scavenging activity (SOSA), nitric oxide radical scavenging activity (NORSA), hydroxyl radical (OH) scavenging activity, lipid peroxidation inhibition capacity (LPIC), total phenolic content (TPC) total flavonoid content (TFC), total carotenoid contents (TCC) and radical scavenging activity as mentioned in (Table 5).

Table 5: Total antioxidant activity of banana peel

Antioxidant activity	Solvents						References	
Antioxidant activity	Water	Chloroform	Acetone	Methanol	Ethanol	Acetyl acetate	n-Hexane	
DPPH	9.8±0.2 ^a	8.7±0.3 ^a	72.83°	28.50±1.30 ^e	19.10 ^c	43.7 ± 0.5^{a}	171.6°	
FRAP	79.28 ^d	16.37±0.38 ^h	ı	$6.98\pm0.34^{\rm f}$	55.10 ^d	-	59.15 ^d	
Metal chelating activity	15.45±0.67 ^a	-	12.3±0.43 ^a	12.20±0.94a	19.4±0.94 ^a	17.35±1.97 ^a	-	
ABTS	69±4.7e	-	75.39±0.59 ^e	35.5±2.4e	28.30±0.85 ^e	-	-	
SOSA	97±4.8e	-	ı	44.7±3.1e	-	-	-	[21, 82, 86-
NORSA	103±4.9a	-	ı	43±1.5 ^e	-	-	-	92]
OH radical scavenging	75.4±2.8 ^e	-	1	39.7±3.7 ^e	-	-	-	
LPIA	-	-	-	89.0ª	88.4ª	3.22±0.85a	-	
TPC	371.74±3.82 ^g	17.85±2.45 ^g		0.54 ± 0.02^{b}	12.58±0.04°	630.14 ± 2.85^{g}	68.83±3.32 ^g	
TFC	8.56±0.22 ^b	15.44±0.19 ^b	16.15±0.28 ^b	51.47±1.94°	18.52±0.06 ^b	- -	-	
TCC	-	-	-	64 ^c	-	-	-	

Note: a = %, b = mg/g, $c = \mu g/g$, $d = \mu M$ Fe²⁺/mg, $e = \mu g/ml$, $f = (mmol Fe^{2+}/100g DW)$, g = GA mg/100 g, h = mg TE/g.

Phenolic compounds

Phenolic compounds are the most essential plant elements that exhibit antioxidant activity by neutralizing lipid free radicals or by preventing hydrogen peroxides from disintegration into free radicals [93, 94]. Vascular plants contain more than 4000 compounds [95]. Phenolic phenolic chemicals are well known for improving quality and nutritional value through modulating color, taste, aroma and flavor in addition to having beneficial impacts on health [96]. Banana peel contains phenolic chemicals, which have been the subject of extensive studies into the effects of dietary polyphenols on human health over the past 20 years. Compared to other fruits, the banana peel contains a high quantity of phenolic. These investigations firmly establish their significance in preventing degenerative disorders, including cancer cardiovascular conditions. investigations firmly establish their significance in preventing degenerative disorders, including cancer and cardiovascular conditions. Several studies have conducted antioxidant effects of polyphenols [97]. As bananas ripened, their overall phenolic content decreased [98]. Some of the health advantages associated with phenolic chemicals include the prevention of cardiovascular disease, cancer, diabetes and obesity [99, 100]. These compounds are effectively used as functional ingredients in the prevention of lipid oxidation [101] and inhibit microbial growth [102]. The phenolic content in green peel is greater up to 15-45 % than in ripe peel whereas it is 52 % higher than that of over-ripened peel [88]. Hang et al. [1] reported that total phenolic content in banana peel ranging from 4.95-47 mg gallic acid equivalent/g dry matter (mg GAE/g DM).

Flavonoid compounds

All plants include a variety of natural compounds called flavonoids, which make up the largest category of plant phenols and more than half of the eight thousand naturally occurring polyphenols [103-105].

Chalcones, isoflavonoids, flavonols, flavanones, flavones, flavanonols anthocyanins are other subcategories of flavonoids. According to published phenolic flavonoid research, and concentrations are natural antioxidant sources [106]. Additionally, flavonoids affect the neurological system and may be used to treat cancer [107]. According to research, the banana peel contains flavones that have enormous potential in the food industry [108]. Banana peel extract contain flavonoid leucocyanidin that promote healing [109].

Antimicrobial activity

Herbal remedies have historically been used to treat a variety of infectious diseases and have been proven beneficial in many cases. Most herbal remedies come from plant components such as leaves, flowers, and stems. To create novel antimicrobial compounds that will operate as a barrier against multidrug-resistant microbes, these extracts may be employed to create new chemical structures and mechanisms of action [110]. Banana peel has been found to contain flavonoids, alkaloids, saponins, triterpenes, tannins. Flavonoid's antibacterial of action mechanisms prevent production of nucleic acids, cytoplasmic membrane function, metabolic activity, adhesion and biofilm generation, altering membrane permeability which reduce the pathogenicity of microbes. Tannins prevent bacterial extracellular enzyme activity and interfere with bacterial metabolism by phosphorylation preventing oxidative [111]. Saponins increase membrane permeability. This rise leads to the cell membrane being unstable and ultimately causing cell hemolysis. Similarly, alkaloids and triterpenoids break down lipid components and increase their permeability, causing the cell membrane to lyse or the cytoplasm of bacterial cells to coagulate (Fig. 3) [112]. As mentioned in (Table 6), previous studies found that banana peel's antibacterial capabilities were efficient against a variety of microbes.

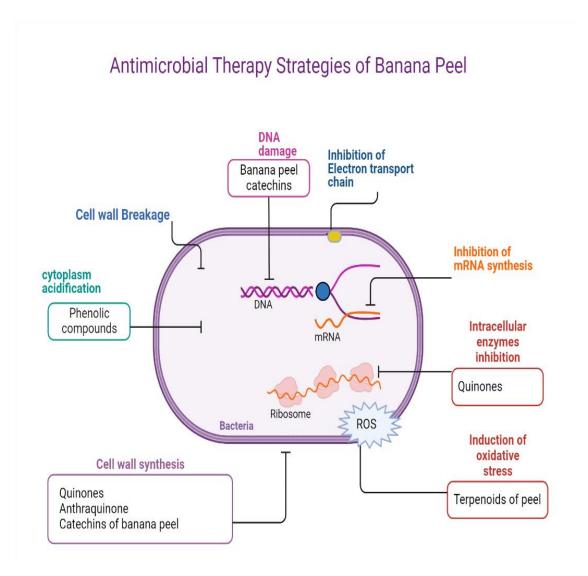


Figure 3: Antimicrobial strategy of banana peel

Table 6: Antibacterial potential of banana peel

Migroongonisms	Extraction solvents Zone of inhibition (mm±standard deviation)						
Microorganisms	Methanol						
			Gram negative			Acetone	7
Pseudomonas aeruginosa	9±0.4	12±0.7	15.86	14±0.9	-	19.57	7
Klebsiella pneumoniae	12±0.4	14±0.6	12±0.5	16±0.5	-	-	7
Serratia marcescens	_	12±0.4	-	9±0.2	11	-	1
Escherichia coli	9±0.3	10±0.3	15.23	12±0.4	-	18.15	
Shigella spp.	-	-	-	-	0	-	
Salmonella typhi	_	12±0.4	10±0.2	9±0.2	-	-	
Salmonella enteritidis	-	10±0.2	-	-	-	-	1
Proteus vulgaris	14±0.3	15±0.3	12±0.2	16±0.4	8	-	7
			Gram positive				
Staphylococcus aureus	_	12±0.4	10±0.2	13±0.3	13	19.57	
Enterococcus faecalis	_	11±0.2	-	12±0.3	-	-	
Aeromonas hydrophila	12±0.3	13±0.3	11±0.2	14±0.3	20	-	[82, 90, 113,
Streptococcus pyogenes	-	10±0.2	-	12±0.3	-	-	114,115]
Listeria monocytogenes	9±0.2	10±0.2	10±0.2	12±0.2	-	-	
Lactobacillus casei	10±0.2	10±0.2	-	12±0.2	-	-	
Bacillus cereus	-	14.17	11.01	-	19.57	-	
Bacillus subtilis	-	10±0.3	15.32	-	-	20.6	
			Fungi				
Aspergillus flavus	-	-	11.76	-	-	17.81	
Aspergillus niger	10±0.2	9±0.2	-	10±0.2	-	-	
Saccharomyces cerevisiae	12±0.2	12±0.2	15.6	20±0.5	-	16.87	
Penicillium citrinum	-	8±0.2	-	9±0.2	-	-	
Candida albicans	-	9±0.2	15.6	14±0.3	-	17.61	7
Alternaria alternata.	-	-	1.37±0.67	-	-	-	

Dermatologist effect

There are different skincare benefits of bananas. There is a certain antimicrobial activity in fruit peels which acts against Staphylococcus and Pseudomonas species [116] as well as high potassium (K) content which is making it the ideal treatment against acne prone-skin. This peel acts as a healing agent by reducing the spread of bacterial skin cells and promoting the healing of pimples and other skin imperfections. It contains antioxidants such as dopamine, ascorbic acid and flavonoids and has anti-ageing benefits [20, 117]. Additionally, it contains vitamin C, which is reported to prevent melanin synthesis, a pigment that causes the skin to darken when exposed to sunlight. Thus, the skin-lightening and soothing properties of banana peel can be associated with antimelanin synthesis and/or degradation because of its anti-inflammatory properties [118]. Potential skin protection, with an skin protection factor (SPF) value 10.67 was demonstrated by the banana peel ethanol extract with bisulphite immersion [119].

Anti-hyperglycemial activity

Defects in insulin secretion, insulin tolerance, or a combination of the following factors known as diabetes mellitus are the most prevalent endocrine diseases [120]. The leading causes of significant health issues in most of the nation [121]. According to a Organization World Health (WHO) report 2013, there are around 347 million diabetic patients worldwide, making it the third leading cause of mortalities [122, 123]. For the treatment of diabetes mellitus, several synthetic oral hypoglycemic drugs, such as biguanides and sulfonylurea, are in addition to insulin. In the prehistoric era of medicine, a variety of plants had been suggested as being beneficial in the identification and management of diabetes mellitus. The peel of different Musa spp. is used to treat and cure diabetes mellitus [124].

It is rich in antioxidants and can boost immunity and lower diabetes risk factors [125]. It has an anti-hyperglycemic effect, which is mostly attributable to the presence of phyto-constituents such as tannins, alkaloids, saponins and flavonoids [120, 126, 127]. Researchers found that anthocyanins, delphinidin, cyanidin and catecholamine are present in banana peel and may also have a role to produce insulin more efficiently from pancreatic beta cells, which in turn has an anti-diabetic impact [8, 128]. Peel has pectin which is one of the key ingredients that has been linked to antihyperglycemic effects. Gallocatechin, quercetin and rutin may have an anti-hyperglycemic effect that has been observed. Gallocatechin promotes peripheral glucose utilization and one of the potential mechanisms for anti-hyperglycemic activity could suppress intestinal glucose absorption or increase insulin generation by pancreatic beta cells. Furthermore, a significant concentration of potassium (K) and sodium (Na) has been linked to the glycemic effect [129]. Chromium (Cr) has been shown to increase glucose tolerance in diabetic elderly individuals and malnourished children. The antihyperglycemic activity was determined using an oral glucose tolerance test and [130] sucrose tolerance test [131].

Anti-ulcerative property

Ulcers are lesions that penetrate the mucosa of the gastrointestinal tract (GIT) [132]. Therefore, the development of the gastroduodenal (peptic) ulcer occurs when the aggressive factors-increased hydrochloric acid (HCl) and pepsin secretion, parietal cell mass and gastrin production-dominate the defensive components. These factors cause auto-digestion and breakdown of gastric mucous. Decreasing stomach production, neutralization of acid pepsin (aggressive factors) and stimulation of protective factors such as mucus and bicarbonate secretion. prostaglandins, mucosal blood flow and nitric oxide are the

major ideas for treating ulcers [133, 134]. Therefore, the main therapy recommendations focus on increasing the synthesis of substances that protect the stomach mucosa in addition to restricting acid secretion, thereby preventing epithelial erosion [135].

Peptic ulcer prevention or treatment is one of the most difficult medical issues since gastric ulcer therapy has downsides and the majority of medications on the market today have limited effectiveness against stomach disorders and are frequently linked to serious adverse effects. In particular, in the treatment of peptic ulcers in various experimental models for the evaluation of anti-ulcer medications, the research of natural products and medicinal plant extracts has emerged as one of the most fascinating and attractive sources of innovative therapy for various gastrointestinal problems. Additionally, they observed the protective effects of the banana peel on gastric mucosa due to its several bioactive ingredients including essential phytochemicals and micronutrients such alkaloids, carotenoids, flavonoids, lignans, tannins. Mechanistic phenolics and characteristics of bioactive components may inhibit ulcer through a number mechanisms, including antioxidant activity, repression of cell growth, activation of apoptosis, repression of cell invasion and intracellular signaling pathways [86].

The remarkable antiulcer activity and cytoprotective properties of the extract of banana peel (*Musa paradisiacal*) on the experimental organisms may be attributed to the phytochemicals such as flavonoids, tannins and saponins [136]. Leucocyanidin, a naturally occurring flavonoid, reportedly preserves the gastrointestinal mucosa from erosions [137]. Leucocyanidin and its synthetic derivatives, hydroxy ethylated leucocyanidin and tetraallyl leucocyanidin, were reported to protect the gastric mucosa in rat models of aspirin-induced erosions by

thickening gastric mucus [138]. Literature indicated that research has been done on laboratory animals (rats and mice). The peel extract has been tested on different acute ulcer models including Alcohol-induced gastric ulcer (AL) in rats [139], Aspirininduced gastric ulcers (ASP) in rats [140] and Pyloric ligation-induced gastric ulcer (PL) in rats [141], Cold-Restrain Stress-Induced Ulcer (CRS) and chronic ulcer model such as Acetic acid-induced ulcer model [142]. Unripe plantain peel extract was found to have a protective effect against (ASP) of stomach mucosa [136]. The peel extract's greatest percentage of ulcer inhibition against ethanol-induced ulcers was 83.33 compared to 100 % against (ASP) and (CRS). Fatimah et al. [143] demonstrated the preventive effect of Musa paradisiaca tepal and skin methanol extracts by demonstrating how they significantly decreased the ulcers.

Anti-inflammatory potential

In response to infection, injury or irritation a process is certain biological called Inflammation **[144]**. The term "inflammation" refers to a complex immunological response that is connected to the gradual release of pro-inflammatory [145]. One cytokines of the main inflammatory mediators is nitric oxide (NO). In addition to NO, other inflammatory mediators such as cyclooxygenase-2 (COX-2), interleukin (IL)-1b, IL-6 and tumor necrosis factor-alpha (TNF-a) are also active are predominantly expressed and inflammatory cell types such as macrophages and mast cells [146, 147]. Therefore, controlling the overproduction of inflammatory mediators, especially proinflammatory cytokines may help to prevent a variety of inflammatory infections [148]. Inflammation appears to be linked to a variety of illnesses, including cancer, allergies, atherosclerosis and even rheumatoid arthritis [149].

products Numerous natural rich phytochemicals including antioxidants and phenolics in fruits and vegetables are the major bioactive components known to show various health benefits [8, 150]. The antiinflammatory actions of numerous plant extract isolated mixtures have already been methodically confirmed, whereas banana peel has been conventionally used as a therapeutic substance for the handling of inflammation [1, 15]. Flavonoid, saponin and tannin contents of banana peel possess antiinflammatory potential [107]. Pathompong et al. [151] indicated that NO inhibitory activity supported the use of Musa spp. peel as medicine for the treatment of inflammation. It indicated that water extract of fresh ripe banana peel exhibits high NO inhibitory activity (IC₅₀ = $6.68\pm0.34 \mu g$).

Conclusion and future prospects

Based on previous information, the banana peel has great biological potential. Concerning nutritional content, the peel can be considered a cheap viable source of dietary fiber because it contains a high amount, which promotes healthy digestion of food. Many studies have proved that banana peel contains high antioxidant and bioactive content that could help the pharmaceutical sector to prevent chronic diseases and increase interest in enhancing food quality. It might provide consumers with a better understanding of the production of valueadded products. On the other hand, several molecular processes, such as metabolomics, genomics and proteomics, enable us in determining the mode of action of several bioactive chemicals to treat a wide range of unclear diseases or infections. Apart from excellent antioxidant sources and high nutritional values for preventing and treating chronic diseases, literature has shown that banana peel may also be used to produce cost-free energy in the form of bioethanol, biodiesel, organic acids, single-cell protein and various important industrial enzymes.

some studies have advocated that peel can be used for skin treatment because of its high antimicrobial potential, which may open a new site for the cosmetic industry. Raw banana peel can be transformed into edible food to avoid its unpleasant taste and increase its consumption.

Authors' contributions

Conceived and designed the experiments: A Akbar & M Hashim, Performed the experiments: M Hashim & Z Hamid, Analysed the data: M Hashim & Z Gul, Contributed materials/ analysis/ tools: A Akbar, Wrote the paper: M Hashim.

References

- 1. Vu HT, Scarlett CJ & Vuong QV (2018). Phenolic compounds within banana peel and their potential uses. A review. *J of Funct Foods* 40: 238-248.
- 2. Jackson T & Badrie N (2003). Utilization of banana (Musa acuminata) peel in wine produced in the Caribbean: Effects on physico-chemical, microbiological and sensory quality of wines. *J of Food Sci and Technol (Mysore)* 40(2): 153-156.
- 3. Ramli S, Alkarkhi AF, Shin Yong Y, Min-Tze L & Easa AM (2009). Effect of banana pulp and peel flour on physicochemical properties and in vitro starch digestibility of yellow alkaline noodles. *Int J of Food Sci and Nutr* 60(sup4): 326-340.
- 4. Sodchit C, Tochampa W, Kongbangkerd T & Singanusong R (2013). Effect of banana peel cellulose as a dietary fiber supplement on baking and sensory qualities of butter cake. *Songklanakarin J of Sci and Technol* 35(6): 641-646.
- 5. Faostat F (2020). Crops. Food and Agric Organ of the UN. Available online.
- 6. Nabi G, Talpur BA, Jarwar IA & Sial M (2020). Exploration of Banana Based Cropping System in District Khairpur Sindh Pakistan.
- 7. Lehmann U & Robin F (2007). Slowly digestible starch—its structure and health implications: a review. *Tr in Food Sci & Technol* 18(7): 346-355.

- 8. Pereira A & Maraschin M (2015). Banana (Musa spp) from peel to pulp: ethnopharmacology, source of bioactive compounds and its relevance for human health. *J of Ethnopharmacol* 160: 149-163.
- 9. Rehman A, Deyuan Z, Hussain I, Iqbal MS, Yang Y & Jingdong L (2018). Prediction of major agricultural fruits production in Pakistan by using an econometric analysis and machine learning technique. *Int J of Fruit Sci* 18(4): 445-461.
- 10. Chai SY, Abbasiliasi S, Lee CK, Ibrahim TAT, Kadkhodaei S, Mohamed MS & Tan JS (2018). Extraction of fresh banana waste juice as non-cellulosic and non-food renewable feedstock for direct lipase production. *Ren Energ* 126: 431-436.
- 11. Guerrero AB, Aguado PL, Sánchez J & Curt MD (2016). GIS-based assessment of banana residual biomass potential for ethanol production and power generation: a case study. *Waste and Biomass Valori* 7(2): 405-415.
- 12. Vu HT, Scarlett CJ & Vuong QV (2017). Optimization of ultrasound-assisted extraction conditions for recovery of phenolic compounds and antioxidant capacity from banana (Musa cavendish) peel. *J of Food Process and Pres* 41(5): e13148.
- 13. Jimenez A, Creissen G, Kular B, Firmin J, Robinson S, Verhoeyen M & Mullineaux P (2002). Changes in oxidative processes and components of the antioxidant system during tomato fruit ripening. *Planta* 214(5): 751-758.
- 14. Davey MW, Stals E, Ngoh-Newilah G, Tomekpe K, Lusty C, Markham R & Keulemans J (2007). Sampling strategies and variability in fruit pulp micronutrient contents of West and Central African bananas and plantains (Musa species). *J of Agric and Food Chem* 55(7): 2633-2644.
- 15. Chabuck ZAG, Al-Charrakh AH, Hindi NKK & Hindi SKK (2013). Antimicrobial effect of aqueous banana peel extract, Iraq. *Res Gate Pharm Sci* 1: 73-75.

- Prasad K, Bharathi K & Srinivasan K (1993). Evaluation of Musa (Paradisiaca Linn. cultivar)-" Puttubale" stem juice for antilithiatic activity in albino rats. *Indian j of Physi and Pharmacol* 37: 337-337.
- 17. Wu H, Xu F, Hao J, Yang Y & Wang X (2015). Antihyperglycemic activity of banana (Musa nana Lour.) peel and its active ingredients in alloxan-induced diabetic mice. Paper presented at the 3rd International Conference on Material, Mechanical and Manufacturing Engineering (IC3ME 2015).
- 18. Zulkifli B, Akmal M, Wahyuni S, Siregar TN & Gholib G (2020). Identification of Active Compounds of Kepok Banana Peel and the Effect on Testosterone Concentration in Male Rats with High-Fat Diet. Paper presented at the E3S Web of Conferences.
- 19. Lal N, Sahu N, Shuirkar G, Jayswal DK & Chack S (2017). Banana: Awesome fruit crop for society.
- 20. Yin X, Quan J & Kanazawa T (2008). Banana prevents plasma oxidative stress in healthy individuals. *Plant Foods for Hum Nut* 63(2): 71-76.
- 21. Vu HT, Scarlett CJ & Vuong QV (2019). Changes of phytochemicals and antioxidant capacity of banana peel during the ripening process; with and without ethylene treatment. *Sci Hortic-Amsterdam* 253: 255-262.
- 22. Englberger L, Schierle J, Marks G C & Fitzgerald MH (2003). Micronesian banana, taro, and other foods: newly recognized sources of provitamin A and other carotenoids. *J of Food Compos and Anal* 16(1): 3-19.
- 23. Morais DR, Rotta EM, Sargi SC, Schmidt EM, Bonafe EG, Eberlin MN & Visentainer JV (2015). Antioxidant activity, phenolics and UPLC–ESI (–)– MS of extracts from different tropical fruits parts and processed peels. *Food Res Int* 77: 392-399.
- 24. Mohd Salim R, Khan Chowdhury AJ, Rayathulhan R, Yunus K & Sarkar MZI (2016). Biosorption of Pb and Cu from

- aqueous solution using banana peel powder. *Desalin and Water Treat* 57(1): 303-314.
- 25. Borhan A, Thangamuthu S, Taha MF & Ramdan AN (2015). Development of activated carbon derived from banana peel for CO2 removal. Paper presented at the AIP Conference Proceedings.
- 26. Karnitz JO, Gurgel LVA, De Melo JCP, Botaro VR, Mel TMS, de Freitas Gil RP & Gil LF (2007). Adsorption of heavy metal ion from aqueous single metal solution by chemically modified sugarcane bagasse. *Bioresou Technol* 98(6): 1291-1297.
- 27. Emaga TH, Andrianaivo RH, Wathelet B, Tchango JT & Paquot M (2007). Effects of the stage of maturation and varieties on the chemical composition of banana and plantain peels. *Food Chem* 103(2): 590-600.
- 28. Hassan P & Peh KK (2018). Chemical Compositions of Banana Peels (Musa sapientum) Fruits cultivated in Malaysia using proximate analysis. *Res J Chem Environ* 22: 108-113.
- 29. Okareh O, Adeolu A & Adepoju O (2015). Proximate and mineral composition of plantain (Musa Paradisiaca) wastes flour; a potential nutrients source in the formulation of animal feeds. *Afr J of Food Sci and Technol* 6(2): 53-57.
- 30. Ozabor P, Ojokoh A, Wahab A & Aramide O (2020). Effect of Fermentation on the Proximate and Antinutrient Composition of Banana Peels. *The Int J of Biotechnol* 9(2): 105-117.
- 31. Durgadevi PKS, Saravanan A & Uma S (2019). Antioxidant potential and antitumour activities of Nendran banana peels in breast cancer cell line. *Indian J of Pharm Sci* 81(3): 464-473.
- 32. Karthikeyan A & Sivakumar N (2010). Citric acid production by Koji fermentation using banana peel as a novel substrate. *Bioresou Technol* 101(14): 5552-5556.
- 33. Emaga TH, Robert C, Ronkart SN, Wathelet B & Paquot M (2008). Dietary

- fibre components and pectin chemical features of peels during ripening in banana and plantain varieties. *Bioresou Technol* 99(10): 4346-4354.
- 34. Davey MW, Van den Bergh I, Markham R, Swennen R & Keulemans J (2009). Genetic variability in Musa fruit provitamin A carotenoids, lutein and mineral micronutrient contents. *Food Chem* 115(3): 806-813.
- 35. Tang Q, Tan P, Ma N & Ma X (2021). Physiological functions of threonine in animals: beyond nutrition metabolism. *Nutr* 13(8): 2592.
- 36. Gorissen SH & Phillips SM (2019). Branched-chain amino acids (leucine, isoleucine, and valine) and skeletal muscle. *Nutr and Skelet Muscle*. pp. 283-298.
- 37. Brosnan ME & Brosnan JT (2020). Histidine metabolism and function. *The J of Nutr* 150(Supplement_1): 2570S-2575S.
- 38. Tall AR & Yvan-Charvet L (2015). Cholesterol, inflammation and innate immunity. *Nat Rev Immunol* 15(2): 104-116.
- 39. Fischetti VA (2008). Bacteriophage lysins as effective antibacterials. *Curr opin in microbiol*. 11(5): 393-400.
- Martínez Y, Li X, Liu G, Bin P, Yan W, Más D, . . . Yin Y (2017). The role of methionine on metabolism, oxidative stress, and diseases. *Amino Acids* 49(12): 2091-2098.
- 41. Kałużna-Czaplińska J, Gątarek P, Chirumbolo S, Chartrand MS & Bjørklund G (2019). How important is tryptophan in human health. *Crit Rev in Food Sci and Nutr* 59(1): 72-88.
- 42. Fernstrom JD & Fernstrom MH (2007). Tyrosine, phenylalanine, and catecholamine synthesis and function in the brain. *The J of Nutr* 137(6): 1539S-1547S.
- 43. Gu C, Mao X, Chen D, Yu B & Yang Q (2019). Isoleucine plays an important role for maintaining immune function. *Curr Protein and Pept Sci* 20(7): 644-651.

- 44. Davis T, Suryawan A, Orellana RA & Fiorotto ML (2010). Leucine acts as a nutrient signal to stimulate protein synthesis. *J of Anim Sci* 88(2).
- 45. Atkins CA, Pate JS & Sharkey PJ (1975). Asparagine metabolism—key to the nitrogen nutrition of developing legume seeds. *Plant Physiol* 56(6): 807-812.
- 46. Nascimento M (2018). Potential uses of arginine in dentistry. *Adv in dent res*. 29(1): 98-103.
- 47. Ros R, Muñoz-Bertomeu J & Krueger S (2014). Serine in plants: biosynthesis, metabolism, and functions. *Tr in Plant Sci* 19(9): 564-569.
- 48. Hall JC (1998). Glycine. *J of Parenter and Enter Nutr* 22(6): 393-398.
- 49. Scott E, Peter F & Sanders J (2007). Biomass in the manufacture of industrial products—the use of proteins and amino acids. *Appl Microbiol and Biotechnol* 75(4): 751-762.
- 50. Stefano M, Marino & Vadim NG (2010). Cysteine function governs its conservation and degeneration and restricts its utilization on protein surfaces. *J of Mol Biol* 404(5): 902-916.
- 51. Tsado A, Okoli N, Jiya A, Gana D, Saidu B, Zubairu R & Salihu I (2021). Proximate, Minerals, and Amino Acid Compositions of Banana and Plantain Peels. *BIOMED Nat and Appl Sci* 1(01): 032-042.
- 52. Muthukumaran J, Srinivasan S, Venkatesan RS, Ramachandran V & Muruganathan U (2013). Syringic acid, a novel natural phenolic acid, normalizes hyperglycemia with special reference to glycoprotein components in experimental diabetic rats. *J of Acute Dis* 2(4): 304-309.
- 53. Hupkens P, Boxma H & Dokter J (1995). Tannic acid as a topical agent in burns: historical considerations and implications for new developments. *Burns* 21(1): 57-61.
- 54. Singh B, Singh JP, Kaur A & Singh N (2016). Bioactive compounds in banana and their associated health benefits—A review. *Food Chem* 206: 1-11.

- 55. Williamson G & Manach C (2005). Bioavailability and bioefficacy of polyphenols in humans. II. Review of 93 intervention studies. *The Am j of Clin Nutr* 81(1): 243S-255S.
- 56. Rasool MK, Sabina EP, Ramya SR, Preety P, Patel S, Mandal N, . . . Samuel J (2010). Hepatoprotective and antioxidant effects of gallic acid in paracetamol-induced liver damage in mice. *J of Pharm and Pharmacol* 62(5): 638-643.
- 57. Garbe D (2000). Cinnamic acid. *Ullmann's Encyclopedia of Industrial Chemistry*.
- 58. Ferguson JW, Tripathi D & Hayes PC (2005). Endoscopic diagnosis, grading and predictors of bleeding in esophageal and gastric varices. *Tech in Gastrointestinal Endoscopy* 7(1): 2-7.
- 59. Perez-Vizcaino F & Duarte J (2010). Flavonols and cardiovascular disease. *Mol Aspects of Med* 31(6): 478-494.
- 60. Kumar N & Pruthi V (2014). Potential applications of ferulic acid from natural sources. *Biotechnol Rep* 4: 86-93.
- 61. Rodrigo-Baños M, Garbayo I, Vílchez C, Bonete MJ & Martínez-Espinosa RM (2015). Carotenoids from Haloarchaea and their potential in biotechnology. *Mar Drugs* 13(9): 5508-5532.
- 62. Nambara E & Marion-Poll A (2005). Abscisic acid biosynthesis and catabolism. *Annu Rev of Plant Biol* 56: 165.
- 63. Young SN (2007). How to increase serotonin in the human brain without drugs. *J of psychiatr & neurosci: JPN* 32(6): 394.
- 64. Wilt T, MacDonald R & Ishani A (1999). -sitosterol for the treatment of benign prostatic hyperplasia: a systematic review. *BJU Int* 83: 976-983.
- 65. Choudhary SP & Tran LS (2011). Phytosterols: perspectives in human nutrition and clinical therapy. *Curr Med Chem* 18(29): 4557-4567.
- 66. Schaller H (2003). The role of sterols in plant growth and development. *Prog in Lipid Res* 42(3): 163-175.

- 67. Khawas P, Das AJ & Deka SC (2017). Banana peels and their prospects for industrial utilization. *Food Processing By-Products and their Utilization* 195-206.
- 68. Kanazawa K & Sakakibara H (2000). High content of dopamine, a strong antioxidant, in cavendish banana. *J of Agric and Food Chem* 48(3): 844-848.
- 69. Riggin RM, McCarthy MJ & Kissinger PT (1976). Identification of salsolinol as a major dopamine metabolite in the banana. *J of Agric and Food Chem* 24(1): 189-191.
- Rebello LPG, Ramos AM, Pertuzatti PB, Barcia MT, Castillo-Muñoz N & Hermosin-Gutierrez I (2014). Flour of banana (Musa AAA) peel as a source of antioxidant phenolic compounds. Food Res Int 55: 397-403.
- 71. Shang C, Gu Y & Koyama T (2021). Major triterpenes, cycloeucalenone and 31-norcyclolaudenone as inhibitors against both α-glucosidase and α-amylase in banana peel. *Int J of Food Sci & Technol* 56(7): 3519-3526.
- 72. Subagio A, Morita N & Sawada S (1996). Carotenoids and their fatty-acid esters in banana peel. *J of Nutr Sci and Vitaminol* 42(6): 553-566.
- 73. Silva A, Morais SM, Falcão M, Vieira I, Ribeiro LM, Viana SM, Teixeira MJ, Barreto FS, Carvalho CA & Cardoso R (2014). Activity of cycloartane-type triterpenes and sterols isolated from Musa paradisiaca fruit peel against Leishmania infantum chagasi. *Phytomed* 21(11): 1419-1423.
- 74. Hassan Z, Jahan IA, Saha GC, Begum F, Nada K & Choudhury J (2010). Studies on the Peel Oil from Two Varieties of Banana. *Bangladesh J of Sci and Ind Res* 45(4): 393-396.
- 75. Walker SE, Shulman KI, Tailor SA & Gardner D (1996). Tyramine content of previously restricted foods in monoamine oxidase inhibitor diets. *J of Clin Psychopharmacol* 16(5): 383-388.
- 76. Lyte M (1997).. Induction of gramnegative bacterial growth by neurochemical containing banana (Musa x

- paradisiaca) extracts. *FEMS Microbiol Lett* 154(2): 245-250.
- 77. Lima GPP, Da Rocha SA, Takaki M, Ramos PRR & Ono EO (2008). Comparison of polyamine, phenol and flavonoid contents in plants grown under conventional and organic methods. *Int J of Food Sci & Technol* 43(10): 1838-1843.
- 78. Nilprapruck P, Meetum P & Chanthasa C (2016). Influence of exogenous spermidine on to peel color change and browning symptom of Musa (AAA group) cv. Kluai Hom Thong at low temperature storage. *Sci, Eng and Health Stud* 38-42.
- 79. Akbar A, Ali I, Samiullah NU, Khan SA, Rehman Z & Rehman SU (2019). Functional, antioxidant, antimicrobial potential and food safety applications of curcuma longa and cuminum cyminum. *Pak J of Bot* 51(3): 1129-1135.
- 80. van der Schaft N, Schoufour JD, Nano J, Kiefte-de Jong JC, Muka T, Sijbrands EJ & Voortman T (2019). Dietary antioxidant capacity and risk of type 2 diabetes mellitus, prediabetes and insulin resistance: the Rotterdam Study. *Eur J of Epidemiol* 34(9): 853-861.
- 81. Kattappagari KK, Teja CR, Kommalapati RK, Poosarla C, Gontu SR & Reddy BVR (2015). Role of antioxidants in facilitating the body functions: A review. *J of Orofac Sci* 7(2): 71.
- 82. Mokbel MS & Hashinaga F (2005). Antibacterial and antioxidant activities of banana (Musa, AAA cv. Cavendish) fruits peel. *Am j of Biochem and Biotechnol* 1(3): 125-131.
- 83. Someya S, Yoshiki Y & Okubo K (2002). Antioxidant compounds from bananas (Musa Cavendish). *Food Chem* 79(3): 351-354.
- 84. Kondo Y, Ohnishi M & Kawaguchi M (1999). Detection of lipid peroxidation catalyzed by chelated iron and measurement of antioxidant activity in wine by a chemiluminescence analyzer. *J of Agric and Food Chem* 47(5): 1781-1785.

- 85. Hertog MG, Kromhout D, Aravanis C, Blackburn H, Buzina R, Fidanza F & Nedeljkovic S (1995). Flavonoid intake and long-term risk of coronary heart disease and cancer in the seven countries study. *Arch of Intern Med* 155(4): 381-386.
- 86. Dahham SS, Mohamad T, Tabana YM & Majid A (2015). Antioxidant activities and anticancer screening of extracts from banana fruit (Musa sapientum). *Acad J of Cancer Res* 8(2): 28-34.
- 87. Bratovcic A, Djapo-Lavic M, Kazazic M & Mehic E (2021). Evaluation of antioxidant capacities of orange, lemon, apple and banana peel extracts by frap and abts methods. *Rev Roum Chim* 66(8-9): 713-717.
- 88. Sundaram S, Anjum S, Dwivedi P & Rai GK (2011). Antioxidant activity and protective effect of banana peel against oxidative hemolysis of human erythrocyte at different stages of ripening. *Appl Biochem and Biotechnol* 164(7): 1192-1206.
- 89. Singh S & Prakash P (2015). Evaluation of antioxidant activity of banana peels (Musa acuminata) extracts using different extraction methods. *Chem Sci Trans* 4(1): 158-160.
- 90. Aboul-Enein AM, Salama ZA, Gaafar AA, Aly HF, Abou-Elella F & Ahmed H (2016). Identification of phenolic compounds from banana peel (*Musa paradaisica* L.) as antioxidant and antimicrobial agents. *J of Chem and Pharm Res* 8(4): 46-55.
- 91. El-Hadary A, El-Shourbagy G, Sulieman A & El-Nemer S (2015). Impact of some technological treatments on antioxidant capacity of banana and potato peel extracts. *Zagazig J of Agric Res* 42(4): 813-824.
- 92. Sulaiman SF, Yusoff NAM, Eldeen IM, Seow EM, Sajak AAB & Ooi KL (2011). Correlation between total phenolic and mineral contents with antioxidant activity of eight Malaysian bananas (Musa sp.). *J of Food Compos and Anal* 24(1): 1-10.

- 93. Maisuthisakul P, Suttajit M & Pongsawatmanit R (2007). Assessment of phenolic content and free radical-scavenging capacity of some Thai indigenous plants. *Food Chem* 100(4): 1409-1418.
- 94. Nasapon P, Ampai P, Maitree S, Chaiyavat C & Pimporn L (2010). Phenolic content and in vitro inhibitory effects on oxidation and protein glycation of some thai medicinal plants.
- 95. Tsamo CVP, Herent MF, Tomekpe K, Emaga TH, Quetin-Leclercq J, Rogez H & Andre CM (2015). Effect of boiling on phenolic profiles determined using HPLC/ESI-LTQ-Orbitrap-MS, physicochemical parameters of six plantain banana cultivars (Musa sp). *J of Food Compos and Anal* 44: 158-169.
- 96. Sengul M, Yildiz H, Gungor N, Cetin B, Eser Z & Ercisli S (2009). Total phenolic content, antioxidant and antimicrobial activities of some medicinal plants. *Pak J of Pharm Sci.* 22(1).
- 97. Scalbert A, Johnson IT & Saltmarsh M (2005). Polyphenols: antioxidants and beyond. *The Am j of Clin Nutr* 81(1): 215S-217S.
- 98. Fatemeh S, Saifullah R, Abbas F & Azhar M (2012). Total phenolics, flavonoids and antioxidant activity of banana pulp and peel flours: influence of variety and stage of ripeness. *Inter Food Res J* 19(3).
- 99. Agbo MO, Uzor PF, Nneji UNA, Odurukwe CUE, Ogbatue UB & Mbaoji EC (2015). Antioxidant, total phenolic and flavonoid content of selected Nigerian medicinal plants. *Dhaka Univ J of Pharm Sci* 14(1): 35-41.
- 100. Boots AW, Haenen GR & Bast A (2008). Health effects of quercetin: from antioxidant to nutraceutical. *Eur J of Pharm* 585(2-3): 325-3.
- 101. Abreu-Naranjo R, Paredes-Moreta JG, Granda-Albuja G, Iturralde G, González-Paramás AM & Alvarez-Suarez JM (2020). Bioactive compounds, phenolic profile, antioxidant capacity and effectiveness against lipid peroxidation of

- cell membranes of Mauritia flexuosa L. fruit extracts from three biomes in the Ecuadorian Amazon. *Heliyon* 6(10): e05211.
- 102. Chaudhry F, Ahmad ML, Hayat Z, Ranjha MMAN, Chaudhry K, Elboughdiri N & Uddin J (2022). Extraction and Evaluation of the Antimicrobial Activity of Polyphenols from Banana Peels Employing Different Extraction Techniques. Separations 9(7): 165.
- 103. Sultana T, Stecher G, Mayer R, Trojer L, Qureshi MN, Abel G & Bonn GK (2008). Quality assessment and quantitative analysis of flavonoids from tea samples of different origins by HPLC-DAD-ESI-MS. *J of Agric and Food Chem* 56(10): 3444-3453.
- 104. Munir H & Sarfraz RA (2014). Medicinal attributes of Aerva javanica native to Pothohar Plateau. *Pak J of Life and Soc Sci* 12(2): 80-86.
- 105. Hazafa A, Rehman K-U, Jahan N & Jabeen Z (2020). The role of polyphenol (flavonoids) compounds in the treatment of cancer cells. *Nutr and Cancer* 72(3): 386-397.
- 106. Akbar A, Gul Z, Aziz S, Sadiq MB, Achakzai JK, Saeed S, Chein SH & Sher H (2022). Bio-Functional Potential and Biochemical Properties of Propelis Collected from Different Regions of Balochistan Province of Pakistan. Oxid Med and Cell Longev 2022.
- 107. Achmad H & Putri AP (2021). Contents of Banana Peel Extract as Hemostasis in Wound Healing. *Ann of the Romanian Soc for Cell Biol* 4800-4810.
- 108. Guerrero-Alva DM (2019). Flavonoids of Organic Banana Peels (*Musa cavendishii*). *Biotechnol* 4(2): 40-45.
- 109. Novak FR, Almeida JAG & Silva RS (2003). Banana peel: a possible source of infection in the treatment of nipple fissures. *J Pediat* 79(221): 6.
- 110. Gul Z Akbar A & Leghari SK (2022). Elucidating Therapeutic and Biological Potential of Berberis baluchistanica

- Ahrendt Bark, Leaf, and Root Extracts. *Front in Microbiol*, 13, 823673-823673.
- 111. Sutanti V & Destyawati AA (2019). The use of yellow kepok banana peel extract (musa paradisiaca L. var bluggoe) as an antibacterial for chronic periodontitis caused by porphyromonas gingivalis. *J of Smart Biopros and Technol P-ISSN* 2686: 0805.
- 112. Brooks G, Carroll K, Butel J, Morse S & Mietzner T (2013). Medical Microbiology. 26th edit: New York: McGraw-Hill.
- 113. Singh CR, Kathiresan K, Boopathy NS, Anandhan S & Govindan T (2013). Evaluation of microbial potential of different coloured banana peels. *Int J Preclin Pharm Res* 4: 62-64.
- 114. Olakunle OO, Joy BD & Irene OJ (2019). Antifungal activity and phytochemical analysis of selected fruit peels. *J of Biol and Med* 3(1): 040-043.
- 115. Saleem M & Saeed MT (2020). Potential application of waste fruit peels (orange, yellow lemon and banana) as wide range natural antimicrobial agent. *J of King Saud Univ-Sci* 32(1): 805-810.
- 116. Alisi C, Nwanyanwu C, Akujobi C & Ibegbulem C (2008). Inhibition of dehydrogenase activity in pathogenic bacteria isolates by aqueous extracts of Musa paradisiaca (Var Sapientum). *African J of Biotechnol* 7(12).
- 117. Vijayakumar S, Presannakumar G & Vijayalakshmi NR (2008). Antioxidant activity of banana flavonoids. *Fitoterapia* 79(4): 279-282.
- 118. Maregesi SM, Kagashe GA & Felix F (2014). Documentation and phytochemical screening of traditional beauty products used in Missenyi District of Tanzania. *J of Cosmet, Dermatol Sci and Appl* 4(05): 355.
- 119. Laeliocattleya R & Wijaya J (2019). The effect of sodium bisulfite immersion to the potential of Candi banana peel ethanol extract as radical scavenger and UV protection. Paper presented at the IOP

- Conference Series: Earth and Environmental Science.
- 120. Tanabe H, Saito H, Kudo A, Machii N, Hirai H, Maimaituxun G, . . . Asahi K (2020). Factors associated with risk of diabetic complications in novel cluster-based diabetes subgroups: a Japanese retrospective cohort study. *J of Clin Med* 9(7): 2083.
- 121. Mbanya JCN, Motala AA, Sobngwi E, Assah FK & Enoru ST (2010). Diabetes in sub-saharan africa. *The lancet* 375(9733): 2254-2266.
- 122. Bharti SK, Krishnan S, Kumar A & Kumar A (2018). Antidiabetic phytoconstituents and their mode of action on metabolic pathways. *Ther Adv in Endocrinol and Metab* 9(3): 81-100.
- 123. Egede LE & Ellis C (2010). Diabetes and depression: global perspectives. *Diabetes Res and Clin Pract* 87(3): 302-312.
- 124. Jain S & Sharma S (1967). Hypoglycemic effect of Musa Sapientum L. flowers. *Planta Med* 4: 439-442.
- 125. Jideani AI, Silungwe H, Takalani T, Omolola AO, Udeh HO & Anyasi TA (2021). Antioxidant-rich natural fruit and vegetable products and human health. *Int J of Food Prop* 24(1): 41-67.
- 126. Sulyman A, Akolade J, Sabiu S, Aladodo R & Muritala H (2016). Antidiabetic potentials of ethanolic extract of Aristolochia ringens (Vahl.) roots. *J of Ethnopharmacol* 182: 122-128.
- 127. Ehiowemwenguan G, Emoghene A & Inetianbor J (2014). Antibacterial and phytochemical analysis of Banana fruit peel. *IOSR J of Pharm* 4(8): 18-25.
- 128. Navghare V & Dhawale S (2017). In vitro antioxidant, hypoglycemic and oral glucose tolerance test of banana peels. *Alexandria j of Med* 53(3): 237-243.
- 129. Rai PK, Jaiswal D, Rai NK, Pandhija S, Rai A & Watal G (2009). Role of glycemic elements of Cynodon dactylon and Musa paradisiaca in diabetes management. *Lasers in Med Sci* 24(5): 761-768.
- 130. Swargiary A, Boro H, Roy MK & Akram M (2021). Phytochemistry and

- Pharmacological Property of Musa balbisiana Colla: A Mini-Review. *Pharmacogn Rev* 15(29).
- 131. Hendra P, Rizki N & Safitri E (2021). Antihyperglycemic activities of Uli banana leaves on oral sugar tolerance. *J of Func Food & Nutr* 2(2): Copyright.
- 132. Ren S, Chen B, Ma Z, Hu H & Xie Y (2021). Polygonum hydropiper extract attenuates ethanol-induced gastric damage through antioxidant and anti-inflammatory pathways. *Brazilian J of Med and Biol Res* 54.
- 133. Ulser T (2016). Antiulcer activity of Musa paradisiaca [banana] tepal and skin extracts in ulcer induced albino mice. *Malaysian j of Anal Sci* 20(5): 1203-1216.
- 134. Suleyman H, Demircan B & Karagoz Y (2007). Anti-inflammatory and side effects of cyclo-oxygenase inhibitors. *Pharmacol Rep* 59(3): 247.
- 135. Chattopadhyay I, Bandyopadhyay U, Biswas K, Maity P & Banerjee RK (2006). Indomethacin inactivates gastric peroxidase to induce reactive-oxygenmediated gastric mucosal injury and curcumin protects it by preventing peroxidase inactivation and scavenging reactive oxygen. *Free Radic Biol and Med* 40(8): 1397-1408.
- 136. Enemchukwu B, Onyedinma E, Ubaoji K & Ngele K (2014). Antiulcer effects of aqueous extract of unripe plantain peels on male wistar (albino) rats. *Biotechnol Indian J* 9(12): 511-515.
- 137. Lewis DA, Fields WN & Shaw GP (1999). A natural flavonoid present in unripe plantain banana pulp (Musa sapientum L. var. paradisiaca) protects the gastric mucosa from aspirin-induced erosions. *J of Ethnopharmacol* 65(3): 283-288.
- 138. Lewis DA & Shaw GP (2001). A natural flavonoid and synthetic analogues protect the gastric mucosa from aspirin-induced erosions. *The J of nutr Biochem* 12(2): 95-100.
- 139. Süleyman H, Akçay F & Altinkaynak K (2002). The effect of nimesulide on the indomethacin-and ethanol-induced gastric

- ulcer in rats. *Pharmacol Res* 45(2): 155-158.
- 140. Goel RK, Das DG & Sanyal AK (1985). Effect of vegetable banana powder on changes induced by ulcerogenic agents in dissolved mucosubstances of gastric juice. *Indian J of gastroenterol: official J of the Indian Soc of Gastroenterol* 4(4): 249-251.
- 141. Shay H (1945). A simple method for the uniform production of gastric ulceration in the rat. *Gastroenterol* 5: 43-45.
- 142. Sarkar S, Sengupta A, Mukhrjee A, Guru A, Patil A, Kandhare AD & Bodhankar SL (2015). Antiulcer potential of morin in acetic acid-induced gastric ulcer via modulation of endogenous biomarkers in laboratory animals. *Pharmacol* 6(7): 273-281.
- 143. Abdullah FC, Rahimi L, Zakaria ZA & Ibrahim AL (2014). Hepatoprotective, antiulcerogenic, cytotoxic and antioxidant activities of Musa acuminata peel and pulp. Novel Plant Bioresources: *Appl in Food, Med and Cosmet* 371-382.
- 144. Wang Q, Kuang H, Su Y, Sun Y, Feng J, Guo R & Chan K (2013). Naturally derived anti-inflammatory compounds from Chinese medicinal plants. *J of Ethnopharmacol* 146(1): 9-39.
- 145. Lin J-Y & Tang C-Y (2008). Strawberry, loquat, mulberry, and bitter melon juices exhibit prophylactic effects on LPS-induced inflammation using murine

- peritoneal macrophages. *Food Chem* 107(4): 1587-1596.
- 146. Kim HP, Son KH, Chang HW & Kang SS (2004). Anti-inflammatory plant flavonoids and cellular action mechanisms. *J of Pharmacol Sci* 0411110005-0411110005.
- 147. McInnes IB & Schett G (2011). The pathogenesis of rheumatoid arthritis. *New Engl J of Med* 365(23): 2205-2219.
- 148. Kim K-M, Kwon Y-G, Chung H-T, Yun Y-G, Pae H-O, Han J-A, . . . Kim Y-M (2003). Methanol extract of Cordyceps pruinosa inhibits in vitro and in vivo inflammatory mediators by suppressing NF-κB activation. *Toxicol and Appl Pharmacol* 190(1): 1-8.
- 149. Devi KP, Malar DS, Nabavi SF, Sureda A, Xiao J, Nabavi SM & Daglia M (2015). Kaempferol and inflammation: From chemistry to medicine. *Pharm res.* 99: 1-10
- 150. Chen S-L, Yu H, Luo H-M, Wu Q, Li C-F & Steinmetz A (2016). Conservation and sustainable use of medicinal plants: problems, progress, and prospects. *Chin Med* 11(1): 1-10.
- 151. Phuaklee P, Ruangnoo S & Itharat A (2012). Anti-inflammatory and antioxidant activities of extracts from Musa sapientum peel. *J of the Med Assoc of Thai Chotmaihet Thangphaet* 95: S142-146.