

Research Article

Pharmacognostic and phytochemical evaluation of *Capparis cartilaginea* Decne., leaf: A medicinal plant

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Abstract

Capparis cartilaginea Decne., a member of the Capparidaceae family, is used in traditional medicine to treat various diseases. However, limited pharmacogenetic and phytochemical studies on its leaves exist. This study aims to perform preliminary phytochemical screening, Pharmacognostic, and physicochemical investigations of *Capparis cartilaginea* leaves. Macroscopy, qualitative and quantitative microscopy of leaves, qualitative phytochemical screening and physicochemical analysis were conducted. Macroscopic analysis showed that the leaves are simple, ovate to elliptic with a retuse apex, hooked spines, and a pubescent surface, changing from dark green to glaucous upon drying. Microscopic examination revealed a stomatal index of 10.90 ± 1.0 for the upper epidermis and 6.01 ± 0.7 for the lower epidermis, with anomocytic stomata. Anatomically, the leaves have a single layered epidermis, palisade parenchyma, and well-developed vascular tissues. The methanol and chloroform extractive values were 5.11% and 5.5%, respectively. The moisture content was 5.065%, and the total ash value was 63.57%, indicating good stability and potential need for purification. Phytochemical screening detected alkaloids, glycosides, phenolics, carbohydrates, amino acids, and proteins in the methanol extract, while glycosides were present in the chloroform extract. Fluorescence analysis revealed multiple phytochemicals, supporting the plant's authenticity. These findings confirm *Capparis cartilaginea* leaves as a valuable resource for Pharmacognostic research and potential therapeutic applications.

Keywords: *Capparis cartilaginea*; Herbal drug; Leaf anatomy; Micromorphology; Pharmacognosy; Phytochemistry

Introduction

Medicinal plants are vital in global healthcare, with 80% of the population relying on herbal remedies, mostly in poor and less developed countries [1-3]. An estimated 50,000 to 80,000 plants are used for medicinal purposes worldwide [4], with traditional medicine prevalent in countries like India, China, and many African nations

[5]. However, lack of documentation and quality control hinders their broader acceptance, especially in developed countries [6].

Standardizing plant materials is essential for ensuring their safety, quality, and efficacy. Pharmacognostic studies, including morphological and anatomical analysis, help authenticate plant species, prevent

adulteration, and guarantee reproducible quality of herbal medicines [7, 8]. These methods are crucial for confirming the authenticity and therapeutic properties of medicinal plants [9, 10].

Capparis cartilaginea Decne., a member of the *Capparis* genus in the Capparidaceae family, is recognized for its significant medicinal and nutritional uses in traditional medicine [11, 12]. The genus *Capparis* comprises approximately 150-200 species found in tropical and subtropical regions across the globe [13, 14], with six species documented in Pakistan [15]. *Capparis cartilaginea* is commonly found in regions such as Southwest and East Africa, Palestine, Iraq, and Pakistan, where it is locally known as "gorilmbuk blatter" in Baluchistan [16] and "Lasaf" in Arabian cultures. This plant contains a range of bioactive compounds, including flavonoids, terpenoids, alkaloids, and sterols, which contribute to its wide-ranging therapeutic properties, such as antibacterial, antioxidant, anti-inflammatory, antifungal, antitumor, insecticidal, and cytotoxic effects [17, 18].

Traditionally, the leaves of *Capparis cartilaginea* have been used to treat various ailments, including itching, shortness of breath, head colds, tumors, wounds, and boils [19]. Studies have also reported its antimicrobial and antioxidant activities [20]. The leaves contain bioactive compounds like carbohydrates, saponins, polyphenols (flavonoids and tannins), triterpenes, sterols, amino acids, and proteins, which are believed to contribute to its medicinal properties [20]. Despite its wide use and promising therapeutic effects, no comprehensive pharmacogenetic study has been conducted on *Capparis cartilaginea*.

Therefore, this study aims to evaluate the pharmacognostic parameters of *Capparis cartilaginea* leaves, including macroscopic, microscopic, physicochemical, phytochemicals and fluorescence

characteristics. This will provide valuable insights into the identification, quality assurance, and standardization of this medicinal plant, supporting its effective use in herbal medicine.

Materials and Methods

Sample collection: The plant was identified by Dr. Uzma Munir, a plant taxonomist. A herbarium sheet was prepared and deposited at the Herbarium of the Centre for Plant Conservation, University of Karachi, Pakistan, to obtain the voucher specimen (G. H. No: 91552).

Macroscopic analysis

Several macroscopic characteristics, including apex, base, color, margin, shape, size, and surface of the leaf samples, were recorded for identification purposes.

Microscopic analysis

Transverse sections (T.S) of fresh leaves were prepared using a sharp razor blade. Fine sections were mounted on a slide with glycerin and observed under a compound microscope (Thomas Scientific, USA). Quantitative microscopic characteristics, such as stomatal type, stomatal index of the upper and lower epidermis, vein islet numbers, and vein termination numbers, were observed.

Preparation of extracts and preliminary qualitative phytochemical screening

The fresh leaves were washed with distilled water, dried in the shade, and then ground into a fine powder. Five grams of the powder were macerated separately in 50 mL of methanol and chloroform solvents and kept in an incubator at 25°C for five days with continuous shaking. The obtained mixture was then filtered using Whatman filter paper No. 4. After evaporation, the extracted values were calculated. The extracts were used for the preliminary qualitative screening of phytochemicals present in the leaves. Various biochemical tests were performed to detect specific phytochemicals, including Mayer's test for alkaloids, Fehling's test for

carbohydrates, Keller-Killiani's test for glycosides, FeCl₃ test for phenolic compounds [21], and Biuret and Ninhydrin tests [22] for proteins and amino acids.

Physicochemical analysis: Physicochemical parameters such as **ash values** (total ash, water soluble ash and acid insoluble ash values) **moisture content** and **fluorescence tests** were performed by standard procedures [23].

Results and Discussion

Morphological analysis of morphological characteristics are essential for pharmacogenetic studies and authenticating the plant in herbal medicine. The results of macroscopic examination of *Capparis cartilaginea* leaf are presented in (Table 1). The shape of the leaf is oval, broad, and fleshy, often terminating in a hooked structure. This hooked apex could serve as a distinguishing feature, potentially providing mechanical support or assisting in the dispersal of the plant. The entire margin and

rounded apex and base of the leaf further contribute to its unique macro morphological profile [15].

Microscopic analysis is essential for plant authentication, quality control, and standardization of plant-based drugs [24, 25]. In *Capparis cartilaginea* leaves, the stomatal index was 10.90 ± 1.0 for the upper epidermis and 6.01 ± 0.7 for the lower epidermis, with anomocytic stomata. The vein islet number ranged from 11 to 14, and the vein termination number was 9 ± 2 (Table 2 & Fig. 1). The higher stomatal index on the upper epidermis suggests more active transpiration on the upper surface, possibly an adaptation to environmental conditions. The anomocytic stomata, characterized by a specific arrangement of subsidiary cells, aid in species identification. Additionally, the vein islet and vein termination numbers provide important information on the leaf's vascular structure, crucial for understanding its transport system [26].

Table 1. Macromorphological characters of *Capparis cartilaginea* Decne., leaf

Characters	Observations
Type	Simple
Apex	Retuse or Emarginate with yellowish brown hooked spine
Base	Obtuse
Color	Dark green after drying glaucous
Margin	Entire or wavy
Shape	Ovate, broad elliptic or orbicular
Size	2-6 X 2-6 cm
Surface	Pubescent

Table 2. Quantitative microscopy of *Capparis cartilaginea* Decne., leaf

S. No.	Parameters	Values
1.	Stomatal index:	
	Upper epidermis	10.90 ± 1.0 (%)
	Lower epidermis	6.01 ± 0.7 (%)
2.	Vein islet number	11-13 (14) mm ⁻²
3.	Vein termination number	9 ± 2 mm ⁻²
4.	Stomata type	Anomocytic

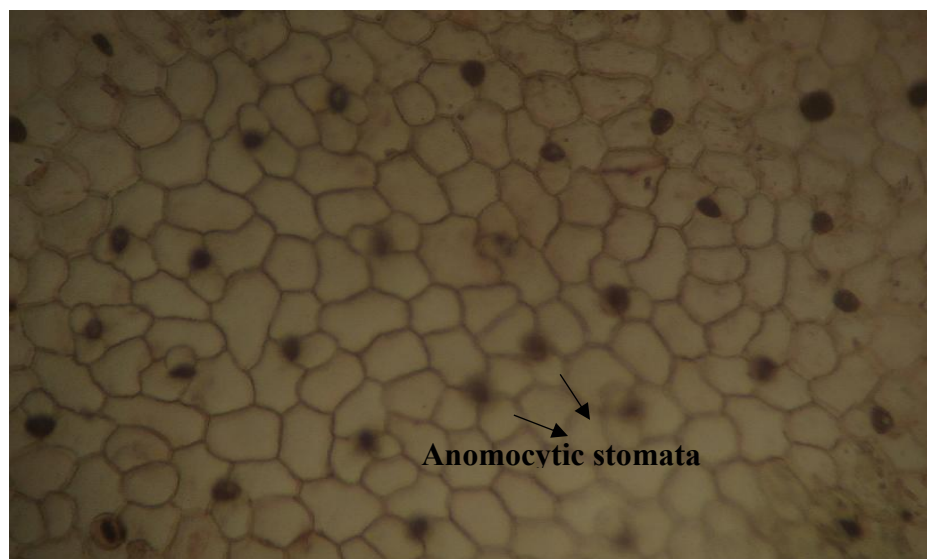


Figure 1. Photomicrograph showing the Transverse Section (T.S) of the leaf lamina across the midrib 100x (Anomocytic stomata)

Leaf anatomy: Transverse section of leaf

Microscopic evaluation is one of the simplest methods used to establish the correct and accurate identity for a plant drug [2]. The transverse section of the *Capparis cartilaginea* leaf revealed a single layered epidermis, palisade parenchyma, spongy parenchyma, xylem, phloem, and a sclerenchymatous pericycle (Fig. 2). The single layered epidermis protects the leaf from water loss and physical damage, while the palisade parenchyma aids in photosynthesis. The spongy parenchyma facilitates gas exchange, and the vascular tissues (xylem and phloem) transport water, nutrients, and sugars, supported by the sclerenchymatous pericycle for structural integrity. The anatomical features observed in the leaf of *Capparis cartilaginea* suggest a well-developed structure that supports both its physiological functions (such as photosynthesis and gas exchange) and its mechanical strength [27]. The anatomical characteristics observed in this study are essential for the identification and quality control of the plant in pharmacognostic evaluations [28].

Extractive values indicate the presence of bioactive compounds in plant material and help assess solvent-specific solubility, which is useful for determining whether a crude drug is exhausted [7, 29]. The extractive values of *Capparis cartilaginea* using methanol and chloroform were 5.11% and 5.5%, respectively (Table 3), suggesting the plant contains moderate levels of both polar (methanol) and non-polar (chloroform) compounds. Methanol extracts polar metabolites, while chloroform extracts non-polar metabolites [30, 31]. These results underscore the plant's potential as a source of diverse bioactive phytochemicals for pharmacological research.

Moisture content affects the stability of crude drugs, with higher levels promoting microbial growth and lower levels improving stability [32]. The air-dried leaf sample of *Capparis cartilaginea* contained 5.065% moisture (Table 3), within the acceptable range of 8-14%, indicating efficient drying and minimal risk of microbial degradation, ensuring safe storage [33-35].

Ash values are key for evaluating the quality, identity, and purity of crude drugs [36, 37,38]. The total ash value of *Capparis*

cartilaginea was 63.57% (Table 3), suggesting significant inorganic content, which could be due to contamination or mineral absorption. The water-soluble ash (4.67%) indicates salts and soluble minerals,

while the acid-insoluble ash (3.85%) reflects silica and non-soluble minerals. These values highlight the need for quality assessment and potential purification [39].

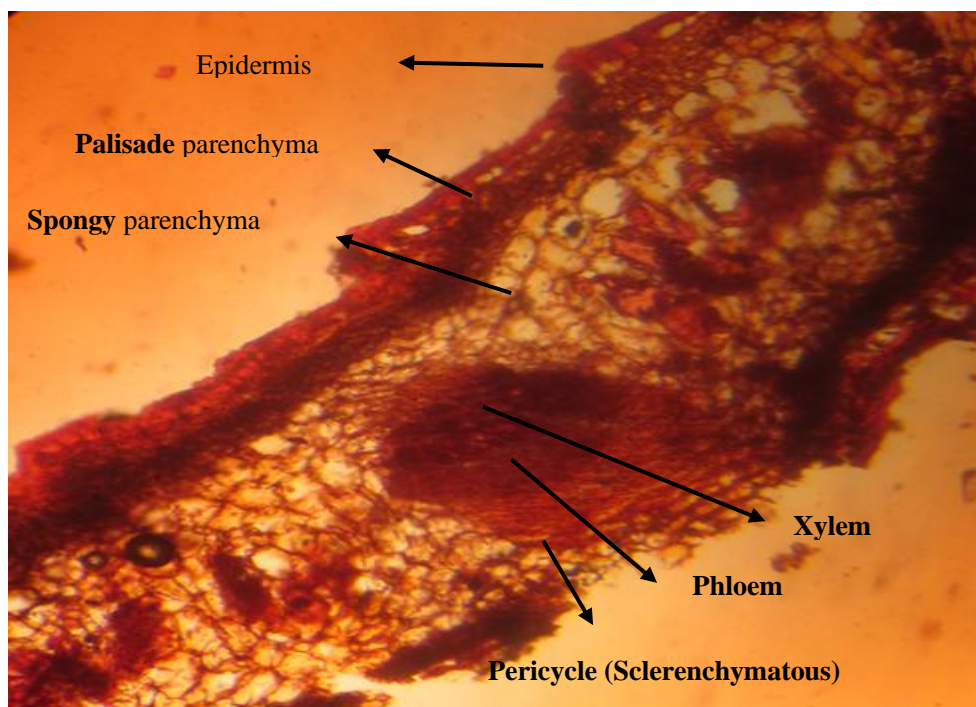


Figure 2. Photomicrograph showing the Transverse Section (T.S.) of the leaf lamina across the midrib 100x

Table 3. Physicochemical analysis of *Capparis cartilaginea* Decne., leaf

S. No.	Quality parameters	% Composition	
		Methanol	Chloroform
1	Extractive Value	5.11±0.73	5.5±0.9
2	Moisture Content	5.065±0.20	
3	Ash value		
	a. Total Ash Value	63.57±0.29	
	b. Water Soluble ash	4.67±0.37	
	c. Acid insoluble Ash	3.85±0.98	

Percentage composition values are presented as Mean ± SEM, n = 3

Phytochemical screening of *Capparis cartilaginea* leaf extract identified bioactive compounds to validate its traditional uses and support future research [40]. The methanolic extract contained alkaloids, glycosides, phenolic compounds, carbohydrates, amino acids, and proteins, while the chloroform extract only contained glycosides (Table 4).

This suggests methanol is more effective for extracting a broader range of bioactive compounds, particularly polar ones [30, 31]. The findings, excluding alkaloids, align with those from Abyan, Yemen [41], with variations likely due to environmental factors [42].

Table 4. Qualitative phytochemical analysis of *Capparis cartilaginea* Decne., leaf

S. No.	Phytochemical constituents	Inference	
		Methanol	Chloroform
1	Alkaloids (Mayer's Test)	+	-
2	Glycosides (Keller Killiani's Test)	+	+
3	Phenolic Compounds (FeCl ₃ Test)	+	-
4	Carbohydrates (Fehling's Test)	+	-
5	Amino acids (Ninhydrin Test)	+	-
6	Proteins (Biuret Test)	+	-

Key: Positive (+) = Present; Negative (-) = Absent

Fluorescence analysis was performed to assess the leaf powder as a crude drug, aiding in plant identification and quality control. Significant color changes were observed under visible and UV light when the dried leaf powder was treated with various solvents

(Table 5). These variations indicate the presence of multiple phytochemicals and suggest the potential of fluorescence analysis in detecting adulteration in crude drug forms [43-45].

Table 5. Fluorescence analysis of dried leaf powder of *Capparis cartilaginea* Decne

S. No.	Reagents	Visible light	UV 366 nm
1	d H ₂ O	Green	Grey
2	75% Ethanol	Light Green	Dark Green
3	1N HCl	Light Brown	Dark Brown
4	1:1 H ₂ SO ₄	Dark Brown	Black
5	5% FeCl ₃	Dark green	Jet black
6	10% NaOH	Yellowish Green	Greyish Green

Conclusion

The comprehensive macroscopic, microscopic, and phytochemical analyses of *Capparis cartilaginea* leave provide valuable insights into its morphological, anatomical, and chemical properties. The distinctive leaf

shape, pubescent surface, and unique apex with hooked spines offer clear identification features. Microscopic parameters, such as the stomatal index, vein structure, and stomata type, contribute to understanding the plant's environmental adaptation and transport

system. Anatomically, the presence of epidermis, palisade parenchyma, and well-developed vascular tissues supports its physiological functions. The extractive values indicate that both polar and non-polar compounds are present, highlighting the plant's potential as a source of bioactive metabolites. Additionally, the low moisture content and high total ash value suggest that the plant is stable for storage and may require purification to ensure purity. Phytochemical screening confirmed the presence of key bioactive compounds, particularly alkaloids and phenolics, which are known for their pharmacological properties. Fluorescence analysis further supports the plant's authenticity and can be used for quality control. These findings underscore the importance of *Capparis cartilaginea* in pharmacognosy and its potential for further therapeutic applications.

Author`s contribution

Conceived, designed and performed the experiments: U Munir, Analyzed the data: S Mansuri, contributed reagents/ materials/ analysis tools: A Perveen, Wrote the paper: I Jamil.

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References

1. WHO, IUCN & WWF (1993). Guidelines on the conservation of medicinal plants, (IUCN) Gland., Switzerland.
2. World Health Organization (2011). Quality control methods for medicinal plants. WHO, Geneva, Switzerland, pp. 9-31.
3. Ekor M (2014). The growing use of herbal medicines: Issues relating to adverse reactions and challenges in monitoring safety. *Front Pharmacol* 4: 177.
4. Chen SL, Yu H, Luo H M, Wu Q, Li CF & Steinmetz A (2016). Conservation and sustainable use of medicinal plants: Problems, progress, and prospects. *Chin Med* 11: 37.
5. Tomlinson TR & Akerele O (2015). Medicinal Plants: Their role in health and biodiversity. University of Pennsylvania Press, Philadelphia.
6. Dahanukar SA, Kulkarni RA & Rege NN (2000). Pharmacology of medicinal plants and natural products. *Indian J Pharmacol* 32: 81-118.
7. Ozarkar KR (2005). Studies on anti-inflammatory effects of two herbs *Cissus quadrangularis* Linn. and *Valeriana wallichii* DC using mouse model. Ph.D. Thesis, University of Mumbai, Mumbai.
8. Sumitra C (2014). Importance of pharmacognostic study of medicinal plants: An overview. *J Pharmacogn Phytochem* 2: 69-73.
9. Nirmal SA, Pal SC & Mandal SC (2012). Pharmacognostic evaluation of *Nyctanthes arbortristis* bark. *Asian Pac J Trop Biomed* 2: 494-500.
10. Kumar A, Verma AK, Gangwar NK & Rahal A (2012). Isolation, characterization and antibiogram of *Mycoplasma bovis* in sheep pneumonia. *Asian J Anim Vet Adv* 7: 149157.
11. Hassanein R, Gabr M, Ahmed A & Hegazi G (2008). Micropropagation of *Capparis cartilaginea* Decne. *Catrina: Int J Environ Sci* 3: 25-32.
12. Bakr RO & El Bishbishy MH (2016). Profile of bioactive compounds of *Capparis spinosa* var. *aegyptiaca* growing in Egypt. *Revista Brasileira de Farmacognosia* 26: 514-520.
13. Rahimifard N, Shojaii A, Mahbobi M, Hafezan G, Bagheri F & Asgarpanah J (2015). Evaluation of antibacterial activity and flavonoid content of two *Capparis* species from Iran. *J Med Plants* 14: 89-94.

14. Coronel E, Alvarenga N, Caballero S & Mereles L (2020). Nutritional and medicinal potential of plant genetic resources of the genus *Capparis* L. and related species in the Gran Chaco. *Rojasiana*, 19: 21-42.
15. Jafri SMH (1974). Capparidaceae. In: Nasir E & Ali SI (Eds.). Flora of Pakistan 71. National Herbarium, Islamabad, pp. 1-42.
16. Lansky EP, Paavilainen HM & Lansky S (2014). Traditional herbal medicines for modern times Caper: The Genus *Capparis*. Boca Raton, FL: CRC Press U.S.A. pp. 39
17. Hamed AR, Abdel-Shafeek KA, Abdel-Azim NS, Ismail SI & Hammouda FM (2007). Chemical investigation of some *Capparis* species growing in Egypt and their antioxidant activity. *J Evid Based Complementary Altern Med* 4: 25-28.
18. Sonbol HS & Al-Balwi ZS (2018). Effect of ethanol extract of *Capparis cartilaginea* on osteoporosis-induced rats. *Int J Pharm Phytopharmacol Res* 8: 59-67.
19. Alsharif B, Babington GA, Radulović N & Boylan F (2022). Volatiles of *Capparis cartilaginea* Decne. from Saudi Arabia. *Plants* 11: 2518.
20. El-Sadek A & Ahmed E (2022). Novel application of *Spirulina platensis* extract as an alternative to the expensive plant growth regulators on *Capparis cartilaginea* (Decne.). *Al-Azhar J Pharma Sci* 66: 29-41.
21. Evans WC (1997). Trease and evans Pharmacognosy. 14th (Eds.). Harcourt brace and company. Asia Pvt. Ltd. Singapore. pp. 343.
22. Gahan PB (1984). Plant histochemistry and cytochemistry: An introduction. Academic Press, Florida, USA. pp.1-123.
23. Evans WC (2002). Trease and evans pharmacognosy. WB Saunders Ltd. London 32, 33, 95-99, 512, 547.
24. Ajaykumar RS & Rajendra DW (2016). Pharmacognostic study and development of quality parameters of *Hamelia patens* jacq. Stems. *Pharm Lett* 8: 6-13.
25. Balasubramaniam G, Sekar M, Badami S (2020). Pharmacognostical, physicochemical and phytochemical evaluation of *Strobilanthes kunthianus* (Acanthaceae). *Pharmacogn J* 12: 731-741.
26. Veeranjanyulu K & Rama VSD (1984). Stomatal frequency and resistance of some tropical members of Asteraceae. *Proc Indian Natl Sci Acad* 50: 317-320.
27. Shaukat M, Huma S, Manyam A, Shahnaz G & Ghazala HR (2010). Pharmacognostic studies on fresh mature leaves of *Holoptetea integritolia* (ROXB) Plarich. *Pax J Bot* 42(6): 3705-3708.
28. Patel S & Zaveri M (2011). Pharmacognostic study of the Roots of *Justica gendarussa* Burm. *J Trad Med* 6: 61-72.
29. Tatiya A, Surana S, Bhavsar S, Patil D & Patil Y (2012). Pharmacognostic and preliminary phytochemical investigation of *Eulophia herbacea* Lindl. Tubers (Orchidaceae). *Asian Pac J Trop Dis* 2: 50-55.
30. Spigno G, Tramelli L & De Faveri DM (2007.) Effects of extraction time, temperature and solvent on concentration and antioxidant activity of grape marc phenolics. *J Food Eng* 81: 200–208.
31. Carrera C, Ruiz-Rodríguez A, Palma M & Barroso CG (2012). Ultrasound assisted extraction of phenolic compounds from grapes. *Anal Chim Acta* 732: 100–104.
32. Okhale SE, Amanabo, MO, Jegede I, Egharevba HO, Muazzam IW & Kunle OF (2010). Phytochemical and pharmacognostic investigation of antidiabetic *Scoparia dulcis* Linn Scrophulariaceae whole plant grown in Nigeria. *Researcher* 2: 7-16.

33. Evans WC (2005). Trease and Evans' Pharmacognosy (Edn15th) Rajkamal Electric press, Delhi, India. pp. 516-536.
34. Adesina GO, Onalapo JA, Ehinmidu JO & Odama LE (2008). Antimicrobial stability of *Alchornea cordifolia* leaf extracts. (Unpublished). International Conference on Research and Development, Ghana.
35. Vinotha S, Ira T & Sri RS (2013). Phytochemical, physicochemical standardization of medicinal plant *Enicostemma Littorale*, Blume. *J Pharm* 3: 52-58.
36. Patnia S & Saha AN (2012). Physicochemical, phytochemical and elemental analysis of stem bark and roots of *Berberis asiatica*. *Adv Appl Sci Res* 3: 3624-8.
37. Mritunjay K, Mondal P, Borah S & Mahato K (2013). Physicochemical evaluation, preliminary phytochemical investigation, fluorescence and TLC analysis of leaves of the plant *Lasia spinose* (Lour) thwaites. *Int J Pharm Pharm Sci* 5: 306-310.
38. Swamy P & Mulla SK (2010). Preliminary pharmacognostical and phytochemical evaluation of *Portulaca quadrifida* Linn". *Int J Pharmtech Res* 2: 699-702.
39. Chandel HS, Pathak AK, Tailang M & Chandra S (2014). Importance of pharmacognostic study of medicinal plants: An overview. *J Pharmacogn Phytochem* 2: 69-73.
40. Ascrizzi R & Cioni PL (2016) Patterns in volatile emission of different aerial parts of caper (*Capparis spinosa* L.). *Chem Biodivers* 13: 904–912.
41. Galib NA & Algfri SK (2016). Phytochemical screening and antioxidant evaluation by DPPH of *Capparis cartilaginea* decne leaves. *J Med Plants* 4: 280-6.
42. Moharram BA, Al-Mahbashi HM, Saif-Ali RIYDH & AliAqlan F (2018). Phytochemical, anti-inflammatory, antioxidant, cytotoxic and antibacterial study of *Capparis cartilaginea* Decne from Yemen. *Int J Pharm Pharm Sci* 10: 38-44.
43. Younus M, Hasan MM, Rehman MS, Abbas K & Sarwar G (2019). Report: Pharmacognostic and physicochemical screening of *Euphorbia nivulia* Buch. Ham. *Pak J Pharm Sci* 32: 1111–1119.
44. Sajid-Ur-Rehman M, Ishtiaq S, Khalil-Ur-Rehman M & Bauer R (2021) Pharmacognostic screening, physicochemical and cytotoxic potential of *Sesuvium sesuvioides* (Fenzyl) Verdc. *Pak J Pharm Sci* 3: 1585–1595.
45. Mondal S & Roy N (2022). Pharmacognostic and phytochemical evaluation of leaf of *Jatropha nana* var. bengalense C.H. Rahaman and S. Mondal: An endemic member of Euphorbiaceae. *Pharmacog Res* 14: 204-10.